HA tag ELISA Kit

Cat.No:AAA23661

Storage：2-8°C.

validity： six months.

The sensitivity of this assay is 1.0 ng/mL

Standard concentration was followed by: 90、30、10、3、1、0 ng/mL.

Both intra-assay CV and inter-assay CV are less than 15%.

The linear range of the kit: 0.05-90 ng/mL.

Accuracy: 95% ± 30%.

FOR RESEARCH USE ONLY; NOT FOR THERAPEUTIC OR DIAGNOSTIC APPLICATIONS!
PLEASE READ THROUGH ENTIRE PROCEDURE BEFORE BEGINNING!

Intended use

This HA TAG ELISA kit is intended Laboratory for Research use only and is not for use in diagnostic or therapeutic procedures.

This kit uses an indirect competitive ELISA method to pre-coat the conjugate antigen on the microbar of the enzyme plate, and the residual HA TAG in the sample and the conjugate antigen pre-coated on the microbar compete with the antibody of HA TAG. After adding the secondary antibody of the enzyme, the TMB substrate is used for color development. The absorbance value of the sample is negatively correlated with the content of HA TAG residue. Multiply by the corresponding dilution to obtain the sample's residual amount of HA TAG.

Sample collection and storage

Plasma

Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 30 minutes at 3000×g at 2-8°C within 30 minutes of collection.

Cell culture supernate and other biological fluids

Remove particulates by centrifugation and assay immediately or aliquot .

Serum

Use a serum separator tube and allow samples to clot for 30 minutes before centrifugation for 10 minutes at approximately 3000×g. Remove serum and assay immediately or aliquot.

Note: The samples should be centrifuged adequately and no hemolysis or granule was allowed. Store samples at -20°C or -80°C. Avoid repeated freeze-thaw cycles.

Materials required but not supplied

1. Standard microplate reader (450nm).
2. Precision pipettes and Disposable pipette tips.
3. 37 °C incubator.

Materials supplied.

Name	96 determinations
Microelisa stripplate	12*8strips
Standard	6 vials
Detection Antibody	6.0 ml*1vial
HRP-Conjugate reagent	11.0ml*1vial
20X Wash solution	50 ml*1vial
Substrate Solution	11 ml*1vial
Stop Solution	6.0 ml*1vial
Closure plate membrane Diluent	2 30ml*1vial
User manual	1

Reagent preparation

1) Bring all kit components and samples to room temperature (20-25°C) for 30 minutes before use.
2) 20×wash solution: Dilute with Distilled or deionized water 1:20.

Assay procedure

1. Prepare all reagents before starting the assay procedure. It is recommended that all. Standards and Samples be added in duplicate to the Microelisa Stripplate.

2. Add standard: Set Standard wells, and testing sample wells. Add standard 100μl to the standard well.

3. Add Sample: Add 100μl sample to the sample well; Blank well doesn't add anything.

4. Add 50 μl DETECTION ANTIBODY to each well except the blank well and mix well. incubate for 45 minutes at 25℃.

5. Aspirate each well and wash, repeating the process four times for a total of five washes. Wash by filling each well with Wash Solution (300μl) using a squirt bottle, manifold dispenser, or auto washer. Complete removal of the liquid

at each step is essential to good performance. After the last wash, remove any remaining Wash Solution by aspirating or decanting. Invert the plate and blot it against clean paper towels.

6. Add 100 µl CONJUGATE to each well except the blank well, mix well. incubate for 30 minutes at 25℃.

7. Aspirate each well and wash, repeating the process four times for a total of five washes. Wash by filling each well with Wash Solution (300µl) using a squirt bottle, manifold dispenser or auto washer. Complete removal of the liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Solution by aspirating or decanting. Invert the plate and blot it against clean paper towels.

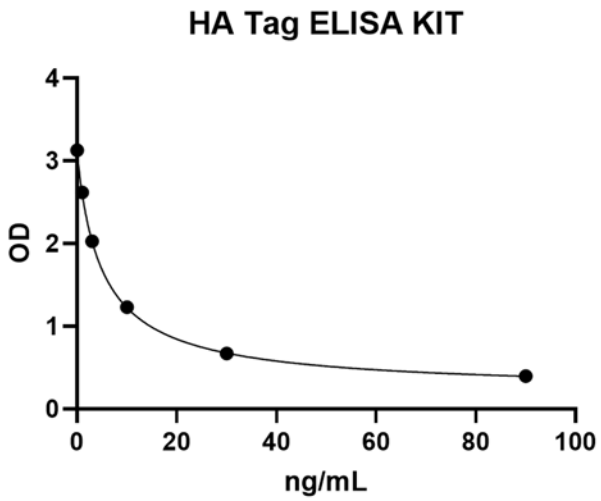
8. Add SUBSTRATE SOLUTION 100µl to each well. Gently mix and incubate for 10 minutes at 25°C. **Protect from light.**

9. Add 50µl Stop Solution to each well.

10. Read the Optical Density (O.D.) at 450 nm using a microtiter plate reader within 15 minutes.

Calculation of results

1) After the detection, the standard concentration was taken as the ordinate, and the corresponding absorbance (OD value) was taken as the abscissa. The four-parameter logistic curve fitting (4-pl) was used to create the standard curve equation, and the concentration value of the sample was calculated by the equation through the absorbance (OD value) of the sample.
2) If the sample is diluted, the final concentration of the sample is determined by. multiplying the concentration value measured by the above method by the dilution multiple.
3) Standard Curve



10. limitations of test methods

1. It is only for scientific research, not for clinical diagnosis.

2. It should be used within the validity period of the kit. Expired products should not be used.

3. It can not be mixed with the kits or components from other manufacturers.

4. Use the sample diluent matched with the kit.

5. If the sample value is higher than the maximum standard concentration value, please dilute the sample properly before re-determination.

6. The test results obtained by other methods are not directly comparable with the test results of this kit.

Product performance index

Four parameters logistic curve (4-pl) was used to fit the model μ g/mL – 8 μ The absolute value of the correlation coefficient (R) of the dose-response curve should not be less than 0.9900 in the range of 0.9900 g / ml.